

Hemato-Oncology Genetic Test Requisition Form

Visit the **UniGen Portal**
to explore our test catalogue
and access your results.



PATIENT INFORMATION

Name Date of Birth / /
Medical Record No. Sex ☐ Female ☐ Male ☐

ORDERING PHYSICIAN

Name Department
Institution e-mail
Phone City, Country Date of Request / /

BILLING INFORMATION

VAT No. Address
Postal Code City Country

SAMPLE INFORMATION

☐ Bone Marrow	☐ Heparine
☐ Blood	☐ EDTA
	☐ PAXgene

Sample collection requirements: *FISH and Karyotype*: collection in lithium heparin or FFPE sample | *NGS*: 2 to 5mL PB/BM in EDTA | *Qualitative/Quantitative tests*: 3 to 5 mL PB/BM in PAXgene or EDTA * | *Clonal tests*: 3 to 5 mL PB/BM in EDTA

* Shipping from Monday to Thursday, never before holidays

☐ FFPE Slides (2 slides per probe, not stained + HE slide with area of interest marked) ☐ FFPE Block ☐ DNA (origin) ☐ Other

Collected at Collection date / / Collection time

CLINICAL INFORMATION

PHYSICIAN'S STATEMENT

I certify that I have obtained the necessary written Informed consent from the patient (or legal representative) to carry out the test(s) I am prescribing here.

I confirm that I have obtained consent for the genetic information to be processed and stored. I confirm that the patient has consented to the retention of any leftover samples, or that I stated in the clinical information that the sample should be discarded.

Physician's signature*

Date (dd/mm/yyyy)*

*REQUIRED FIELDS



LYMPHOID NEOPLASIAS

Chromosome analysis - Karyotype: ☐ non-stimulated cultures

CLL

FISH:

- ☐ (13q-, 11q-, 17p-, +12) - OncoFISH
- ☐ (13q-, 11q-, 17p-, +12, IGH) - OncoFISH
- ☐ ATM del(11)(q22.3)
- ☐ BCL2 rea. (18q21.3)
- ☐ del(13)(q14.3) (D13S319)
- ☐ IGH rea. (14q32)
- ☐ TP53 del(17)(p13.1)
- ☐ Trisomy 12

Molecular analysis:

- ☐ IGVH hypermutation status
- ☐ TP53 gene (somatic mutations)

ALL

FISH:

- ☐ BCR-ABL t(9;22)
- ☐ IGH rea. (14q32)
- ☐ KMT2A rea. (11q23.3)
- ☐ PDGFRβ rea. (5q32)

Molecular analysis:

- ☐ t(9;22)(BCR-ABL) – RT-PCR
- ☐ t(1;19)(E2A-PBX1) – RT-PCR
- ☐ t(12;21)(TEL/AML1) - RT-PCR
- ☐ t(4;11)(KMT2A/AF4) - RT-PCR
- ☐ BCR-ABL p190 (E1A2) quantification
- ☐ BCR-ABL p210 (B2A2, B3A2) quantification
- ☐ E2A-PBX1 quantification
- ☐ TEL/AML1 quantification
- ☐ KMT2A/AF4 quantification
- ☐ BCR-ABL (p190) fusion gene sequencing
- ☐ BCR-ABL (p210) fusion gene sequencing

☐ Other

MM

FISH:

- ☐ (13q, 17p, 1q21.3, IGH) - OncoFISH
- ☐ (1q, 17p, t(4;14), t(14;16)) - OncoFISH
- ☐ C-MYC rea. (8q24.21)
- ☐ del(13)(q14.2) (RB1)
- ☐ del/amp CDKN2C/CKS1B (1p32.3/1q21-22)
- ☐ IGH rea. (14q32)
- ☐ IGH-CCND1 t(11;14)
- ☐ IGH-FGFR3 t(4;14)
- ☐ IGH-MAF t(14;16)

Molecular analysis:

- ☐ TP53 gene (somatic mutations)

Lymphomas

FISH:

- ☐ ALK rea. del(2p); t(2;5)
- ☐ API2-MALT1 t(11;18)
- ☐ BCL6 rea. (3q27-28)
- ☐ C-MYC rea. (8q24.21)
- ☐ IGH rea. (14q32)
- ☐ IGH-BCL2 t(14;18)
- ☐ IGH-MALT1 t(14;18)

Molecular analysis:

- ☐ B cells (IGH) clonal rearrangement
- ☐ IGK clonal rearrangement
- ☐ TCRB clonal rearrangement
- ☐ TCRD clonal rearrangement
- ☐ TCRG clonal rearrangement
- ☐ BRAF gene (V600 mutation)
- ☐ MYD88 (p.Leu265Pro)

MYELOID NEOPLASIAS

Chromosome analysis - Karyotype: ☐ non-stimulated cultures

AML

FISH:

- ☐ (t(15;17), inv(16), t(8;21), KMT2A, 5q-, 7q-, MECOM) - OncoFISH
- ☐ CBFB (inv(16)/t(16;16))
- ☐ del(20)(q12)
- ☐ del(7)(q22q31)
- ☐ EGR1 del 5(q31)
- ☐ KMT2A rea. (11q23.3)
- ☐ PML-RARα t(15;17)
- ☐ RUNX1-RUNX1T1 t(8;21)
- ☐ RPN1-MECOM (inv/t(3))

Molecular analysis:

- ☐ Myeloid Neoplasias Panel
- ☐ t(9;22)(BCR-ABL) - RT-PCR
- ☐ t(15;17)(PML-RARα) - RT-PCR
- ☐ t(8;21) (AML1/ETO) - RT-PCR
- ☐ inv(16)/t(16;16)(CBFB-MYH11) - RT-PCR
- ☐ BCR-ABL p190 (E1A2) quantification
- ☐ BCR-ABL p210 (B2A2, B3A2) quantification
- ☐ PML-RARα (BCR1/2) quantification
- ☐ PML-RARα (BCR3) quantification
- ☐ AML1-ETO quantification
- ☐ CBFB-MYH11 (A) quantification
- ☐ CBFB-MYH11 (E/D) quantification
- ☐ KIT gene (exons 8-11, 13, 17 e 18)
- ☐ NPM1 gene (exon 12)
- ☐ FLT3 gene (ITD; TKD mutations)
- ☐ IDH1/IDH2 genes (variants of exon 4)

☐ Other

MPN

FISH:

- ☐ BCR-ABL t(9;22)
- ☐ FGFR1 rea. (8p11.2)
- ☐ PDGFRα rea. (4q12)
- ☐ PDGFRβ rea. (5q32)

Molecular analysis:

- ☐ JAK2 (V617F; exons 12-15), CALR (exon 9), MPL (exon 10) (somatic mutations)
- ☐ t(5;12) (TEL-PDGFRβ) - RT-PCR
- ☐ JAK2 gene (V617F mutation)
- ☐ KIT gene (D816V)

MDS and MDS/MPS

FISH:

- ☐ (5q-, 7q-, 20q-) - OncoFISH
- ☐ (5q-, 7q-, 20q-, cen8) - OncoFISH
- ☐ del(7)(q22q31)
- ☐ del(20)(q12)
- ☐ EGR1 del(5)(q31)
- ☐ TP53 del(17)(p13.1)
- ☐ Trisomy 8

Molecular analysis:

- ☐ Myeloid Neoplasias Panel
- ☐ UBA1 (codon 41)

CML

FISH:

- ☐ BCR-ABL t(9;22)

Molecular analysis:

- ☐ t(9;22)(BCR-ABL) - RT-PCR
- ☐ BCR-ABL p190 (E1A2) quantification
- ☐ BCR-ABL p210 (B2A2, B3A2) quantification
- ☐ BCR-ABL (p190) fusion gene sequencing
- ☐ BCR-ABL (p210) fusion gene sequencing

NOTICE

1. General description, purpose and risks

The purpose of a genetic test is to examine DNA and look for any specific changes, or variations that could be related to certain diseases or conditions, and the risk of developing certain disorders and conditions.

This test might not provide conclusive results about genetic status. While some genetic variants are known in scientific databases to be pathogenic and others are known to be benign, some of the genetic variants detected may be of uncertain significance.

In addition, current limitations in genetic testing technologies and/or in scientific/medical knowledge may not identify relevant variants that are associated with a disease or symptoms.

However, such results do not rule out the possibility of a genetic disease or predisposition to such a disease. Further testing are recommended in such cases.

2. Use of data

In order to carry out the test, Unilabs CGC - Centro de Genética Clínica e Patologia S.A. will collect, process and use personal data relating to health. The policy, followed in accordance with the laws in force, is described at www.unilabs.pt.